Antimelanoma Effect

Article

In Vitro and In Ovo Assessment of Betulinic Acid

Dorina Coricovac^{1,2}, Iulia Pînzaru^{1,2}, Ștefana Avram^{2,3,*}, Ioana Macașoi^{1,2}, Codruta Soica^{2,4} and Cristina Dehelean^{1,2}

¹ Department of Toxicology and Drug Industry, Faculty of Pharmacy, "Victor Babes" University of Medicine and Pharmacy, Eftimie Murgu Sq. no. 2, 300041 Timişoara, Romania; dorinacoricovac@umft.ro (D.C.); iuliapinzaru@umft.ro (I.P.); macasoi.ioana@umft.ro (I.M.); cadehelean@umft.ro (C.D.) ² Research Center for Pharmaco-Toxicological Evaluations, Faculty of Pharmacy, "Victor Babes" University of

Medicine and Pharmacy, Eftimie Murgu Sq. no. 2, 300041 Timişoara, Romania ³ Department of Pharmacognosy, Faculty of Pharmacy, "Victor Babes" University of Medicine and Pharmacy, Eftimie Murgu Sq. no. 2, 300041 Timişoara, Romania; stefana.avram@umft.ro

⁴ Department of Pharmaceutical Chemistry, Faculty of Pharmacy, "Victor Babes" University of Medicine and Pharmacy, Eftimie Murgu Sq. no. 2, 300041 Timişoara, Romania; codrutasoica@umft.ro

* Correspondence: stefana.avram@umft.ro; Tel.: +40-722-542-528

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Abstract: Although it recorded a breakthrough for the treatment of metastatic melanoma in the last decade, this disease remains a challenge in terms of finding an efficient treatment, reducing secondary resistance to treatment, and understanding the complexity of the molecular mechanisms involved in its development, progression and metastasis. This study aims to verify the multitarget effect of a natural compound, betulinic acid (BA), a pentacyclic triterpene, as antimelanoma agent by applying two experimental models: a human melanoma cell line—A375 and the chick chorioallantoic membrane model. The methods applied in this study were: MTT cell viability assay for cytotoxicity assessment and the chorioallantoic membrane assay (CAM) for antiangiogenic evaluation. The results indicated a significant decrease of A375 cells viability after a 72 h BA treatment even at the lowest concentration tested—1 μ M (61.95% viable cells), with a calculated IC50 of 9.437 µM. In addition, BA inhibited not only the in ovo A375-induced tumor growth but also the angiogenesis on the primary site at 72 h post application. These data highlight the potential antimelanoma effect of BA by targeting the tumor cells via multiples pathways as inducing cell death and suppressing the angiogenic process, a must have for tumor development.

Keywords: melanoma; betulinic acid; A375 melanoma cell line; angiogenesis; cytotoxicity

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Introduction

Cutaneous melanoma is one of the most common type of melanoma that origins from the malignant transformation of the epidermal melanocytes (cells that generate melanin pigment) induced by ultraviolet radiation (UV)'s deleterious effects [1,2]. According to the report elaborated by the National Cancer Institute



Surveillance, Epidemiology, and End Results Program (NCI-SEER) in 2019, melanoma represents the fifth most common type of cancer in men and women worldwide [3]. It is well-known that metastatic melanoma patients present a reduced rate of tumor regression, develop resistance to targeted treatment and have a low survival rate, features that could be related to the immunological character of this disease that exhibits a high degree of heterogeneity at histological and clinical level and an augmented number of mutations [2,4–6]. The complexity of melanoma, particularly when metastasis occurs, makes it exceedingly difficult to treat. In the last decade, the treatment of metastatic melanoma has known a great development by the introduction of immunotherapy such as inhibitors of BRAF, CTLA4 (cytotoxic T-lymphocyte antigen-4) and PDI (programmed death-1) with promising results [1,2,5,6]. Still, these compounds were associated with multiple immune adverse reactions and another inconvenient is represented by the fact that not all patients can tolerate this kind of treatment. Besides the increased rate of response to single or combined immunotherapy, the durability of the response could be considered a variable in the light of melanoma cells' ability to develop drug resistance [1,2,5]. Novel strategies are required to enhance the effectiveness of the existent treatment and to overcome the potential drawbacks.

A hallmark of cancer, and particularly of solid tumors including melanoma, is represented by angiogenesis also known as tumor neovascularization that implies the formation of novel blood vessels from an existent vascular system [7-9]. Tumor-associated angiogenesis requires a constant activation due to the high demand of the uncontrollable proliferation tumor cells, whereas in physiological conditions (as wound healing) is an incidental process, and it is considered a pivotal factor due to its involvement in the formation of primary tumor, tumor survival, and in the development of metastases [7,8,10]. In cutaneous melanoma, angiogenesis is vital for the transition from radial to vertical growth phase and could be used as an indicator of the aggressiveness of the tumor, a highly vascularized tumor being correlated with poor prognosis [8,9]. The existing knowledge regarding tumor-associated angiogenesis converts this process into an important therapeutic target, significant steps being performed in this direction with the novel antiangiogenic agents, but in the case of malignant melanoma, the response to antiangiogenic therapy is rather low. A possible explanation is the vascular mimicry (melanoma cells invade the endothelial cellular network and simulate the endothelial cell functions) or the capacity of melanoma cells to evade blood stream and to grow along the existent blood vessels [10]. This different behavior of melanoma cells in terms of angiogenesis as compared to other solid tumors makes it exceedingly difficult to treat and finding new therapeutic approaches becomes mandatory.

A potential therapeutic alternative for the treatment of melanoma could be considered betulinic acid (BA), a natural molecule, member of the triterpenes pentacyclic family, that attracted an increased interest in recent years due to its broad spectrum of biological effects as antitumor, antiangiogenic, anti-inflammatory, immunomodulatory, antiviral, antidiabetic, antihyperlipidemic, hepatoprotective, etc. [II–I5]. The potent anticancer effect proved against multiple tumor cells of different origin (melanoma, breast cancer, neuroblastoma, glioblastoma, lung cancer, colorectal carcinoma, prostate carcinoma, hepatocellular carcinoma, pancreatic cancer, etc.) [I3] together with the very low/absent degree of toxicity exerted in normal cells make this compound a reliable candidate as anticancer agent [II]. Significant insights were taken into the underlying mechanism of BA antitumor activity: (i) inductor of apoptosis via mitochondrial intrinsic pathway; (ii) regulator of cell cycle and angiogenic pathway by suppressing the expression of the specificity protein—Sp transcription factors—SpI, Sp3 and Sp4, of EGFR—epidermal growth factor receptor, cyclin DI and VEGF expression; (iii) modulator of NF-KB activity, and (iv) antimetastatic effect via inhibition of epithelial-to-mesenchymal transition [II,13,16]; still, it is far from being completely elucidated. In the light of the facts stated above, this study aims to verify the effect of betulinic acid (BA) as an antimelanoma agent *in vitro* by exerting a multitarget potential: cytotoxic in human (A₃₇₅) melanoma cell line and antiangiogenic on an *in ovo* melanoma model.

Materials and Methods

Reagents

The reagents used in the study were of analytical-grade purity and suitable for cell culture conditions. Betulinic acid (purity (HPLC) \geq 98%), dimethyl sulfoxide (DMSO—purity (GC) \geq 99.9%), phosphate saline buffer (PBS), Trypan blue (purity (HPLC) \geq 80%), trypsin-EDTA solution, MTT cell proliferation kit I (code: 11465007001, Roche) were purchased from Sigma Aldrich, Merck KGaA. The cell culture medium—Dulbecco's Modified Eagle's Medium (DMEM) and the other reagents used for cell culture—fetal bovine serum (PBS), antibiotics mixture, were bought from ATCC and Thermo Fisher Scientific (Waltham, MA, USA).

Cell Lines

To perform the experiments assigned for the present study was used a human melanoma cell line—A375 (ATCC^{*} CRL-1619TM) acquired as frozen item from American Type Culture Collection (ATCC). This cell line is an immortalized cell line, with adherent growth properties, epithelial morphology (a detailed characterization of the cell line can be found on the manufacturer site)), and classified as Biosafety level I (present minimal potential risk both to laboratory personnel and the environment) [17].

Cell Culture

A375 melanoma cells require the following cell growth conditions: a specific culture medium—Dulbecco's Modified Eagle's Medium (DMEM—ATCC^{*} 30-2002TM) high glucose—4500 mg/L enriched with 10% fetal bovine serum (FBS) and 1% mixture of antibiotics (Penicillin and Streptomycin) to avoid cells' contamination. Through the experiments, the cells were incubated in standard conditions (at 37 °C and 5% CO₂) and cultured according to the manufacturers' recommendations. The cells were counted automatically by the means of CountessTM II Automated Cell Counter and Trypan blue.

Cell Viability Determination

The cytotoxic effect of BA on A375 cells was assessed by the means of a standard cell viability assay—MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide) that is applied to measure the metabolic activity of the cells as a marker of cell viability and cytotoxicity. The experimental protocol applied consisted of the following steps: (1) cells' seeding (10⁴ cells/200 μ L medium/well) in 96-well plates overnight; (2) stimulation of cells with increasing concentrations of BA in DMSO (1, 5, 10, 25, 50 and 75 μ M) in fresh culture medium for 72 h; (3) at 72 h post-treatment it was added 10 μ L MTT reagent/well and incubated for 3 h at 37 °C, followed by addition of 100 μ L of solubilization buffer/well and maintenance at room temperature and dark for 30 minutes; and (4) absorbance values measurement at 570 nm using a microplate reader (xMarkTMMicroplate, Bio-Rad, CA, USA). The obtained data are expressed as the mean % of viable cells compared to the control \pm SD (each concentration was tested in triplicate). The control cells were considered the cells stimulated with the solvent—DMSO.

Microscopical Evaluation of Melanoma Cells Morphology

The potential changes in melanoma cells morphology induced by BA treatment for 72 h were monitored and photographed under bright light illumination of the Olympus IX73 inverted microscope (Olympus, Tokyo, Japan) equipped with a DP74 camera.

In Ovo Melanoma Model Using the Chorioallantoic Membrane (CAM) Assay

In order to verify the impact of BA on melanoma angiogenesis process it was obtained an *in ovo* model of human melanoma using A375 melanoma cells and CAM assay.

For this experimental part, white chicken eggs acquired from local providers were used. The *in ovo* melanoma model was performed according to the experimental protocol described by the literature and adapted to our laboratory conditions with slight modifications [18,19]. In brief, the basic protocol consisted of the following steps: (1) egg disinfection with ethanol and incubation at 37 °C and controlled humidity; (2) aspiration of approximately 5 mL of albumen (day 3 of incubation); (3) opening of resealable windows on the upper side of the eggs (day 4 of incubation); (4) inoculation of A375 cells (10⁵ cells/3 µL culture medium) on the 10th day of incubation inside plastic rings disposed on the CAMs.

On the following day (0 h), test (10mM BA in DMSO) and control (1% DMSO) solutions were applied inside the rings on top of the CAMs containing melanoma cells. Daily, 3 µL of samples were applied onto the developing specimens. *In ovo* stereomicroscopic (Discovery 8 Stereomicroscope, Zeiss, Göttingen, Germany) observation, after 72 and 96 h, focused on changes in the tumor-associated vascular response next to alteration of tumor development. Relevant images were captured using the Axio CAM 105 color, Zeiss digital camera and processed by Zeiss ZEN software, ImageJ and GIMP. All experiments were performed in triplicate.

Statistical Analysis

The statistical analysis of the obtained results was performed using Graph Pad Prism 8 software. One-way ANOVA statistical test was applied to define the statistical differences between the BA and DMSO-treated cells followed by Dunnett's post-test (**** p < 0.0001). The data are expressed as mean values \pm standard deviation (SD).

Results

BA Treatment for 72 h Triggered a Marked Cell Death in A375 Melanoma Cells

Treatment of A375 melanoma cells with BA in DMSO solution (1, 5, 10, 25, 50 and 75 μ M) for 72 h led to a concentration-dependent decrease of cells' viability (Figure 1). A statistically significant reduction of cells viability percentage (61.95%) was observed even at the lowest concentration tested—1 μ M, the values of viability percentage at the highest concentration—75 μ M being lower than 20%. The calculated IC50 was 9.437 μ M.

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Figure 1. *In vitro* assessment of BA impact (I, 5, 10, 25, 50 and 75 μ M) on viability of A375 cells at 72 h post-stimulation by the MTT cell viability assay. The results are expressed as cell viability percentage (%) normalized to DMSO-stimulated (the solvent used for BA) cells. The data represent the mean values \pm SD of three independent experiments performed in triplicate. One-way ANOVA analysis was applied to determine the statistical differences compared with the control (DMSO) group, followed by Dunnett's multiple comparisons post-test (**** *p* < 0.0001).

BA Treatment for 72 h Induced Morphological Changes in A375 Melanoma Cells

The cytotoxic effect of a compound can be also evaluated by the changes induced in cellular morphology. Following this line, were monitored microscopically the morphological changes induced by BA treatment in A375 cells after 72 h. As it can be noticed in Figure 2, BA induced significant changes in cells' shape, adherence to the plate and confluency in a concentration-dependent manner. At the lowest concentration tested—1 μ M were observed round cells floating in the culture medium, still the confluency was not markedly affected. Starting with 25 μ M, BA induced significant changes in cells morphology as round shape, extremely low adherence to the plate and a reduced confluency, and at the highest concentration—75 μ M cellular debris was predominant. DMSO treatment induced morphological changes only at the highest concentrations—50 and 75 μ M as round cells that floated within the culture medium, but the confluency was not markedly affected as in the case of BA-treated cells. These data confirm the cell viability results.



Figure 2. Microscopical aspect of A₃₇₅ human melanoma cells after BA (1, 5, 10, 25, 50 and 75 μ M) and DMSO treatment for 72 h. The pictures were taken using the 10x objective.

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BA Effects on Melanoma Angiogenesis In Ovo

We investigated *in ovo* the influence of BA on tumoral angiogenic process, including on A375 melanoma cells development. The evaluation was performed compared to the solvent control, diluted DMSO. No signs of toxicity were noted up to 72 h after treatment, in neither test nor control applied samples. The viability of the specimens was reduced after this period, registering a higher death rate of the specimens at 96 h post treatment.

In the first days after application, BA did not influence tumor angiogenesis at an important degree, showing a spokes wheel aspect outside the ring, and especially towards the migrating tumor tissue, but not inside the ring. At 72 and 96 h, BA induced a reduction of the primary tumor mass development, and a limited vascular reaction (Figure 3). In contrast, a limited effect on the forming secondary tumor areas could be observed. BA did not influence the invasiveness of tumor cells, and secondary tumors were observed. Still, when BA was applied directly on secondary formed tumors, melanoma growth was reduced. BA exerted an inhibitory effect both on A375 cells growth and angiogenesis at the primary site after 72 h post application, while allowing the formation of secondary tumors, well vascularized peritumoral but not intratumorally, compared to the control specimens.



Figure 3. BA treatment impact on A₃₇₅ melanoma cell graft and tumor formation on chick embryo chorioallantoic membrane (CAM) by the means of stereomicroscopy.

Discussion

The intricacy of melanoma as concerns the cellular, biochemical, genetic and immunological events underlying its development, progression and metastasis sets this pathology among the most challenging type of cancer in terms of treatment [4,6]. Even if the results of the latest strategies applied for the treatment of metastatic melanoma are promising, there is still a major drawback considering the well-known capacity of melanoma cells to evade cell death and acquire resistance to treatment. Multiple directions of research were initiated in the field of metastatic melanoma treatment including the assessment of natural compounds for

anticancer properties that gained an increased interest due to the multiple pharmacological effects exhibited and the reduced adverse effects.

The present study was developed to verify the antimelanoma properties of BA, a pentacyclic triterpene with lupan skeleton, by assessing its cytotoxic potential in a human melanoma cell line—A375 and its ability to suppress tumor angiogenesis, a key process in melanoma progression and metastasis. The A375 melanoma cell line was selected as model for the *in vitro* cytotoxicity assessments based on A375 cells ability to keep the features of the human genitor and to present B-RAF and CDKN2 mutations, specific for cutaneous melanoma [18].

Betulinic acid ($_{3}\beta$ -Hydroxy-lup-20(29)-en-28-oic acid), a bioactive compound of natural origin, has been subjected to multiple pharmacological studies in the last two decades; the first study acknowledging the antimelanoma activity of BA was conducted by Pisha et al. which began in 1995 [20]. Further studies proved the *in vitro* antimelanoma properties of BA, not only as single agent [16,21–26] but also in combination with gamma irradiation [21] or vincristine [27]. The cytotoxicity results observed after BA treatment in melanoma cells described in the abovementioned studies [16,21,22] support our results. Besides the antimelanoma effect, BA also showed potent anticancer activity against multiple tumor cells, as: breast cancer, glioblastoma, lung cancer, colorectal carcinoma, prostate carcinoma, pancreatic cancer, neuroblastoma, etc. [12,13,28].

Our results indicate a cytotoxic effect of BA dependent on concentration, but based on the data obtained in our previous study [16] that showed a marked cytotoxic effect of BA in A375 cells only at 50 μ M after 24 h, it could be stated that the effect is also time dependent (1 μ M BA at 72 h reduced the A375 percentage cells viability at 61.95%—see Figure 1). Suresh and collaborators also proved a dose dependent cytotoxic effect of BA in A375 cells after 24 and 48 h stimulation with an IC50 value of 154 μ M [29]. Treatment of A375 cells with BA solution (0–100 μ M) for 96 h determined a marked cytotoxicity and an IC50 of 13.3 μ M [30], a value significantly lower as compared to the value obtained by Suresh et al. after 24 and 48 h (IC50 = 154 μ M), which indicates the BA induces a time-dependent cytotoxicity. These data are in agreement with our results.

The chick chorioallantoic membrane (CAM) assay is frequently applied as a pre-screening experimental model for the study of cancer (in terms of tumor aggressiveness, angiogenesis, efficacy of a novel chemotherapeutic agent) that can be applied not only for melanoma, but also for other types of cancer like thyroid cancer [18,31]. This method presents several advantages as compared to mouse models, such as the development of the primary tumor requiring a shorter time, reduced costs, reliable results, multiple evaluations concurrently, and does not require ethical approval since, according to the EU, it is considered as a non-animal method that respects the 3R criteria (replace, reduce, refine) [18,31].

To the best of our knowledge, the findings regarding the antiangiogenic effect of BA in melanoma are rather missing, but there is described the antiangiogenic mechanism of action in breast cancer [27,28], prostate cancer [32,33], and pancreatic cancer [34]. An antiangiogenic effect of BA was also shown in human endometrial adenocarcinoma cells by suppressing prolidase, HIF-1 α and VEGF expressions [35], and in human colorectal cells by down-regulating VEGF expression [36].

Our results indicate that treatment with BA impaired the in ovo tumor growth. Concerning the melanoma-associated angiogenesis, BA proved an inhibitory effect but only on the primary tumor angiogenesis (see Figure 3). The antiangiogenic potential of BA was previously assessed by our group concerning the effect on normal angiogenesis as a preliminary evaluation for tumor treatment application. Direct antiangiogenic effect on endothelial cells next to rapid vessel maturation was described *in ovo* [37].

Conclusions

The present study has shown that BA induced a concentration-dependent cytotoxicity in A375 melanoma cells by reducing cells' viability and triggering morphological changes. In addition, BA exhibited an inhibitory effect on the *in ovo* tumor growth and suppressed the angiogenesis associated to the primary tumor. These results represent the basis for further mechanistic elucidation of BA-suppressed angiogenesis and to characterize its anti-invasive and antimetastatic potential.

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